

3D PRINTING OF A MICROFLUIDIC DEVICE FOR THE PREPARATION OF LIPIDIC NANOPARTICLES

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Fused deposition modeling (FDM) 3D printing

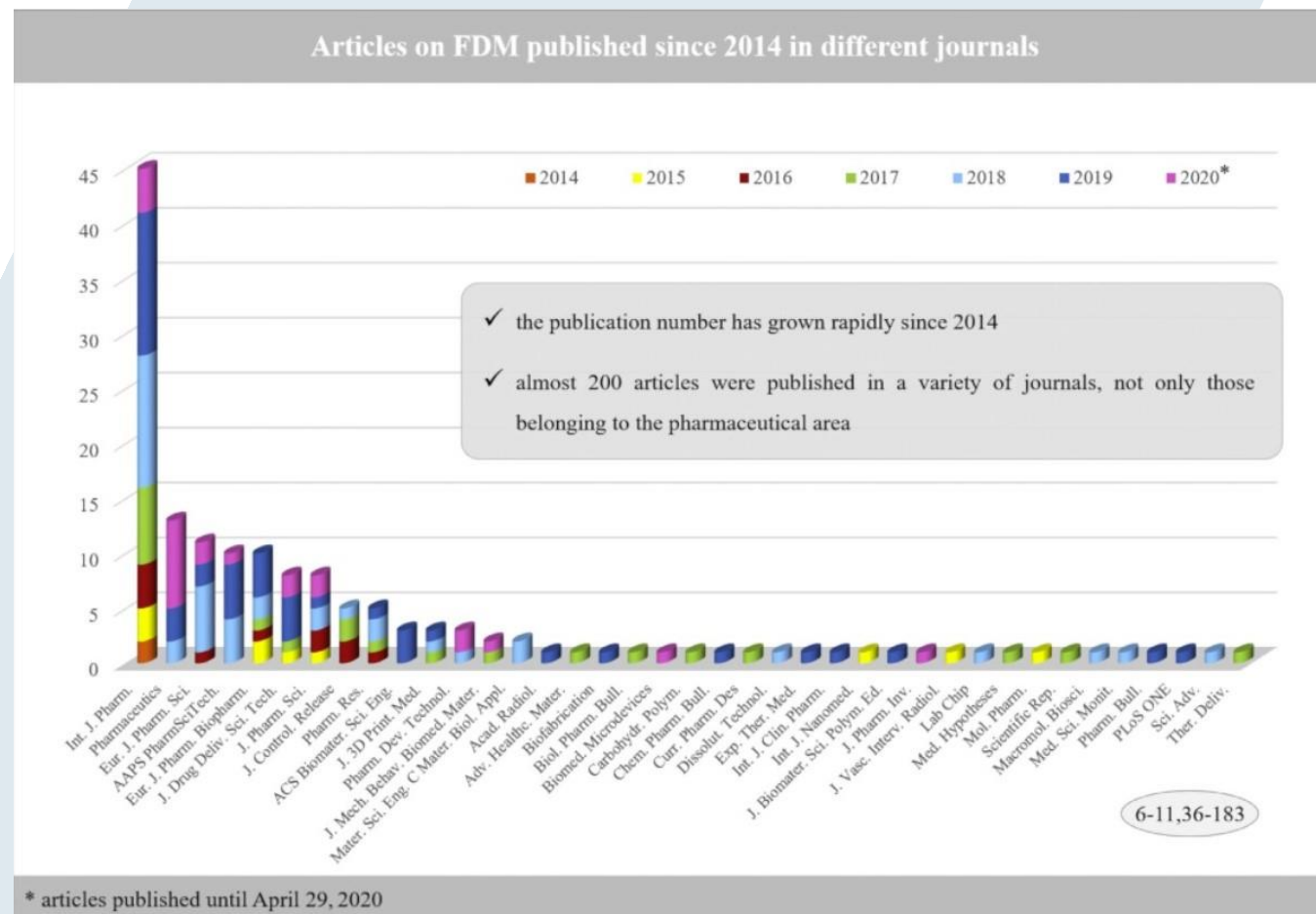


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- 3D printing technology that allow to print objects layer by layer
- Increasing interest on its application in the pharmaceutical field during the last ten years
- Production of pharmaceutical devices (patches, microneedles, stents, films, etc)
- Direct production of pharmaceutical forms (tablets, suppositories)
- Personalized medicine (Form, dosage, etc)

But it is not only direct production of pharmaceutical forms or devices....



© Makerbot

A. Melocchi, M. Uboldi, M. Cerea, A. Foppoli, A. Maroni, S. Moutaharrik, L. Palugan, L. Zema, A. Gazzaniga, A Graphical Review on the Escalation of Fused Deposition Modeling (FDM) 3D Printing in the Pharmaceutical Field, J. Pharm. Sci. (2020). <https://doi.org/10.1016/j.xphs.2020.07.011>.

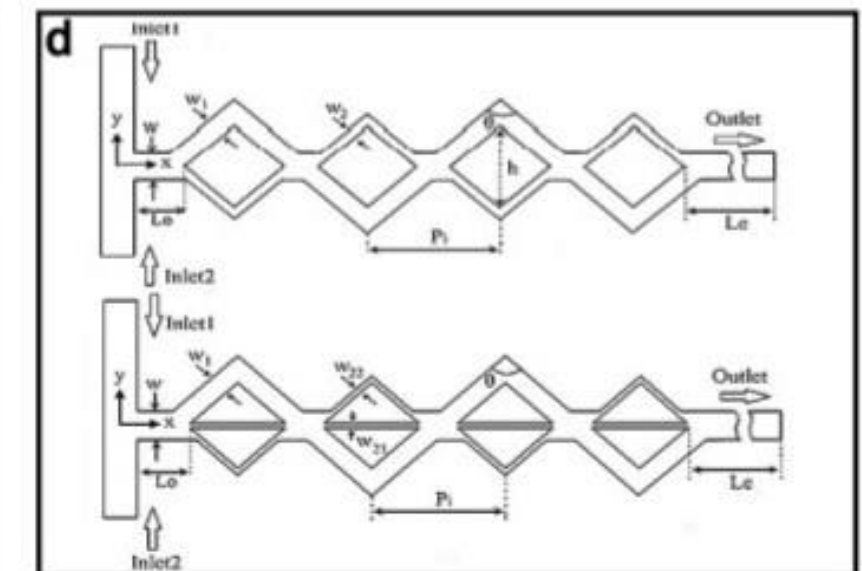
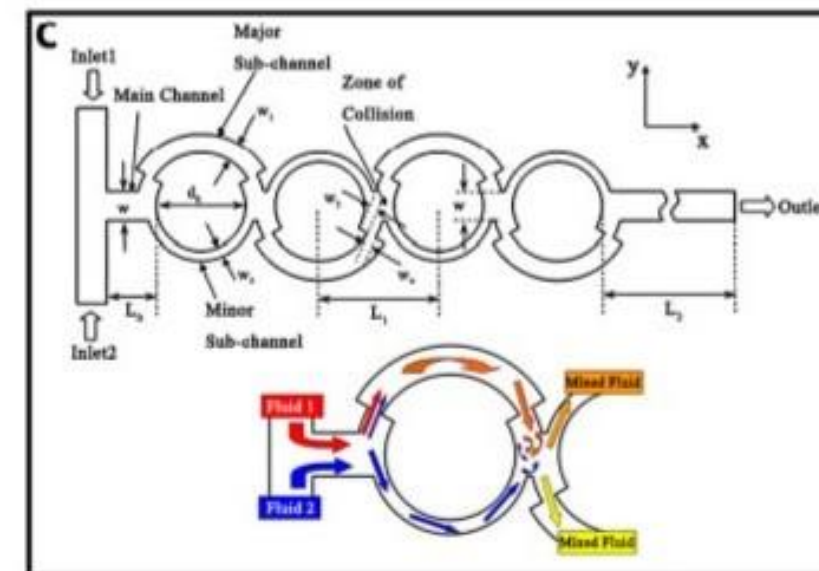
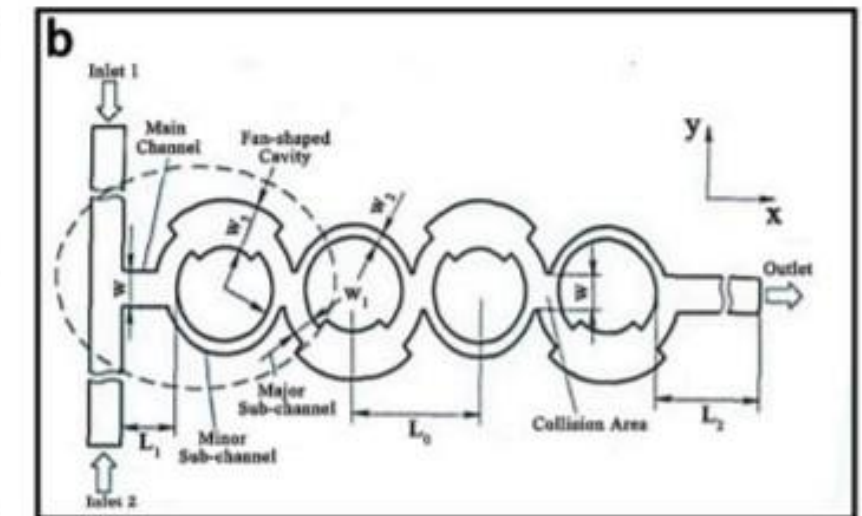
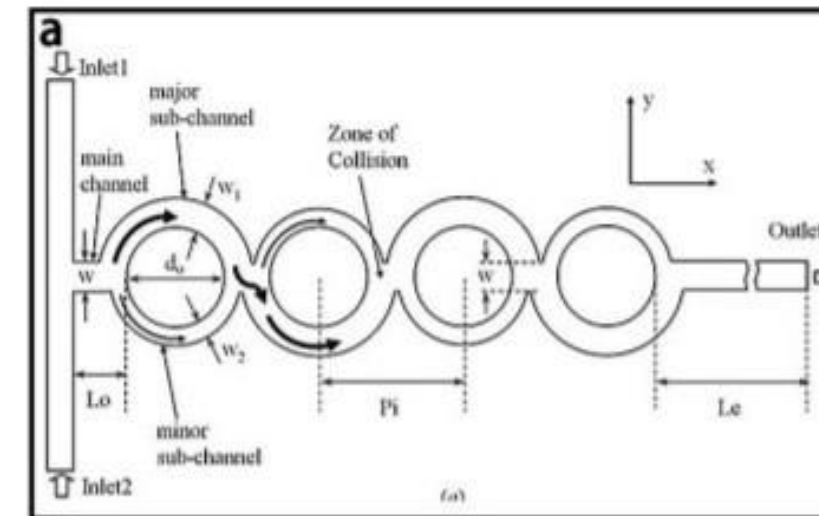
Microfluidics



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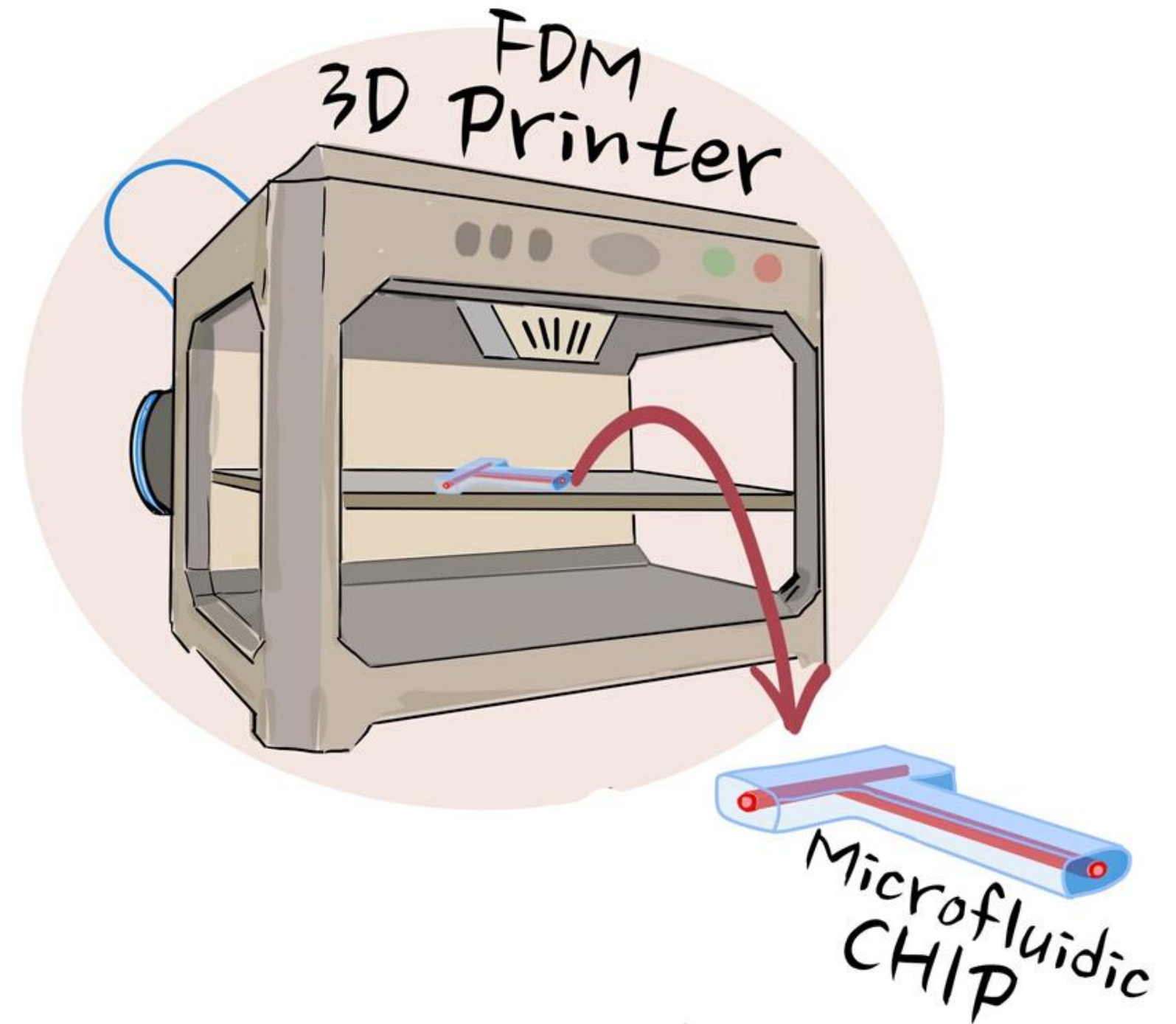


- Controllable and scalable technique for the production of nano or micro particles
- Precise control of micromixing under laminar flow
- One-step production
- High production rates
- Higher drug encapsulation compared to conventional techniques



Why is convenient to produce 3D printed microfluidic chips?

- Device design by CAD software
- Material choice
- Low cost
- Complete personalization of the device
- Easy to produce
- Good resolution

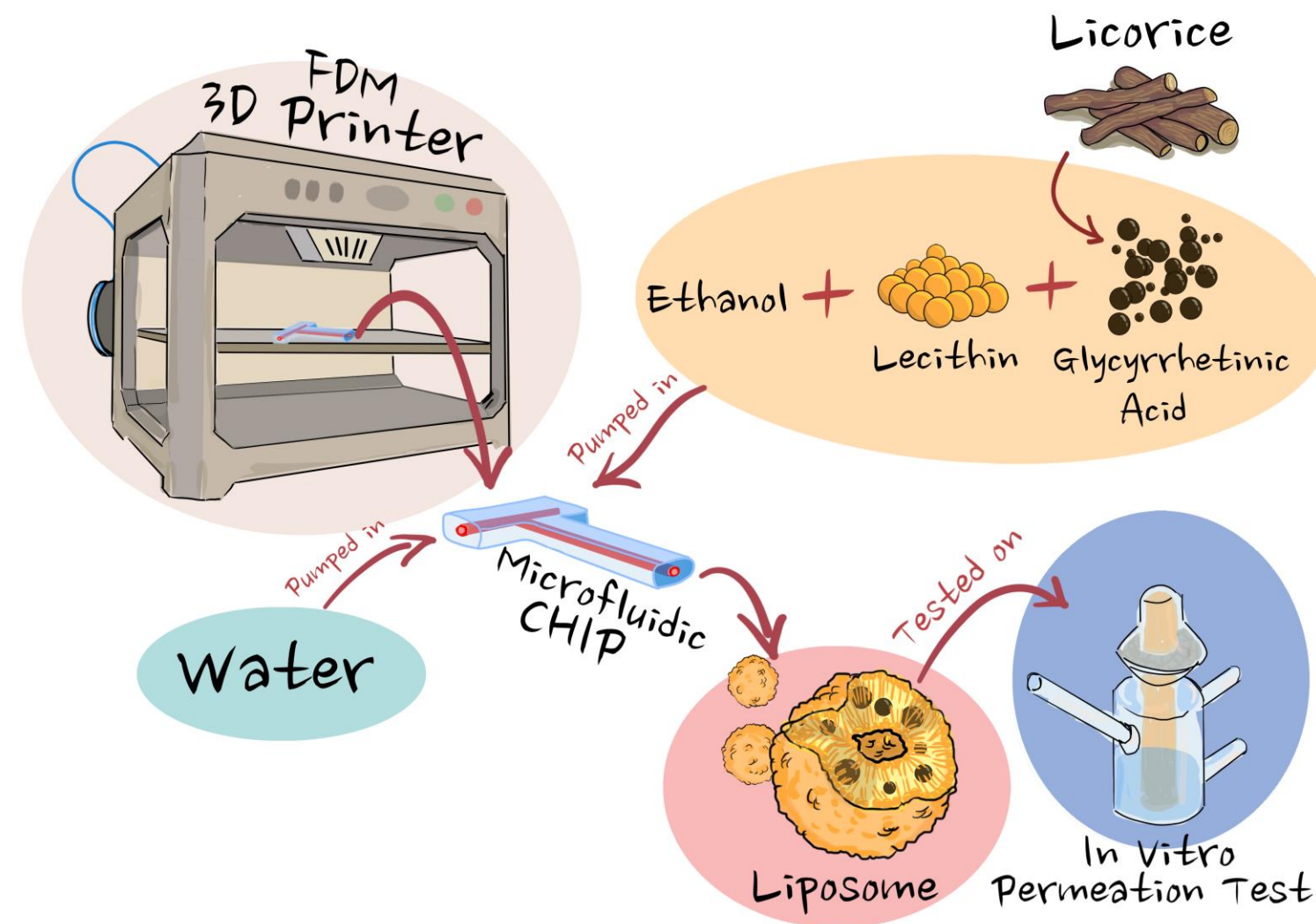


A case study

3D-printed microfluidic chip for the preparation of glycyrrhethinic acid-loaded ethanolic liposomes

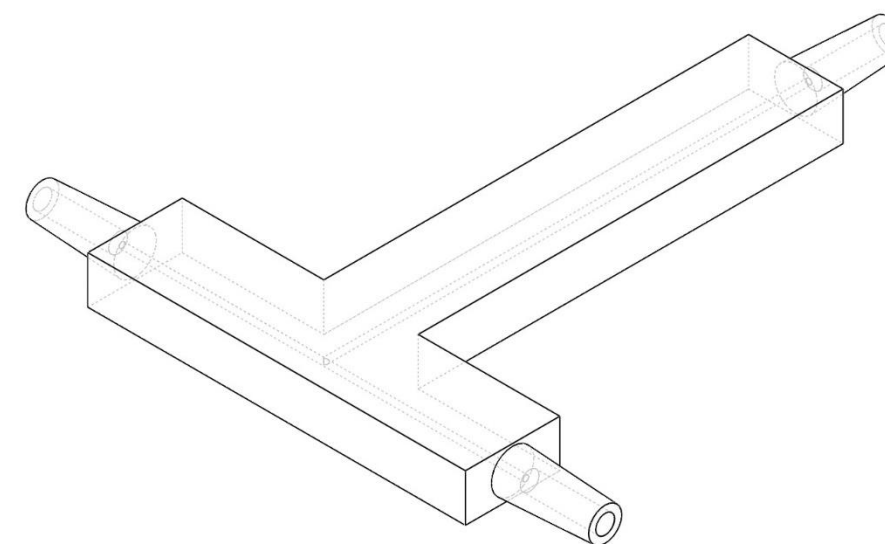
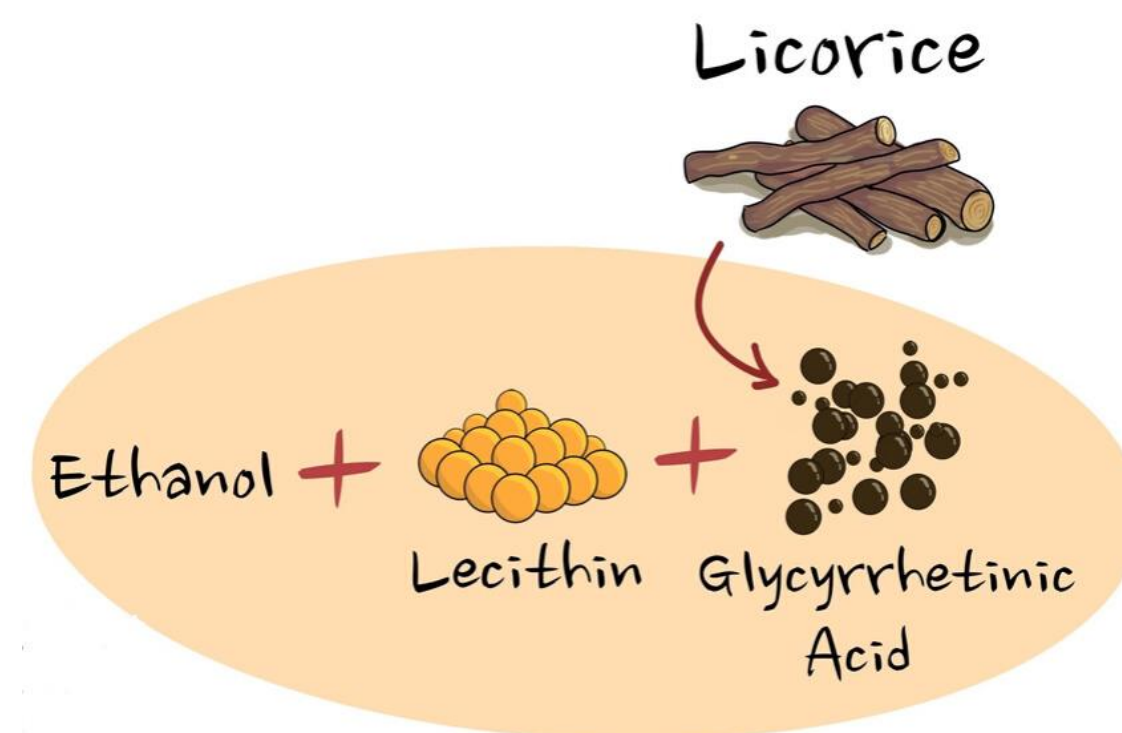


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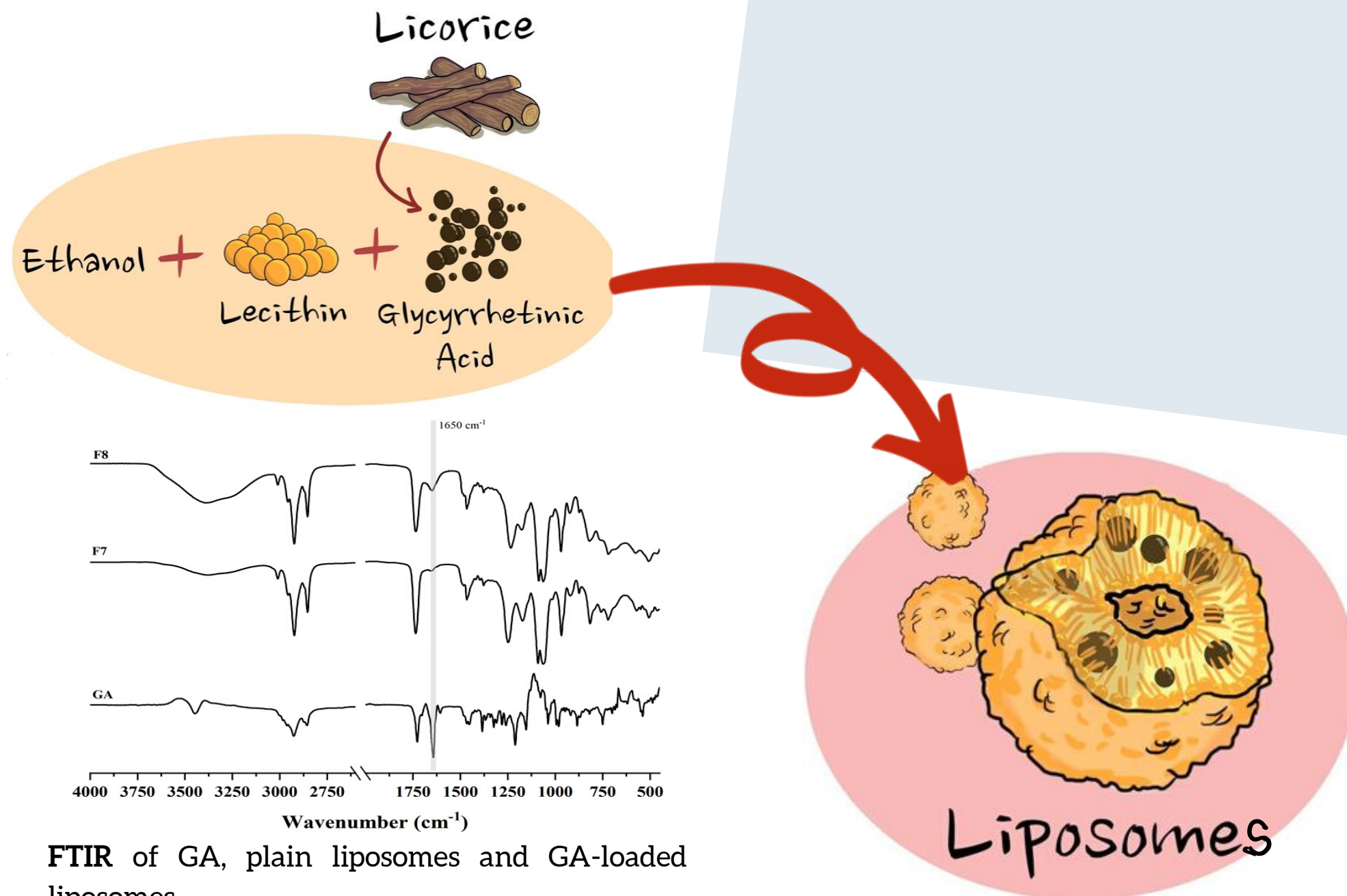


BACKGROUND

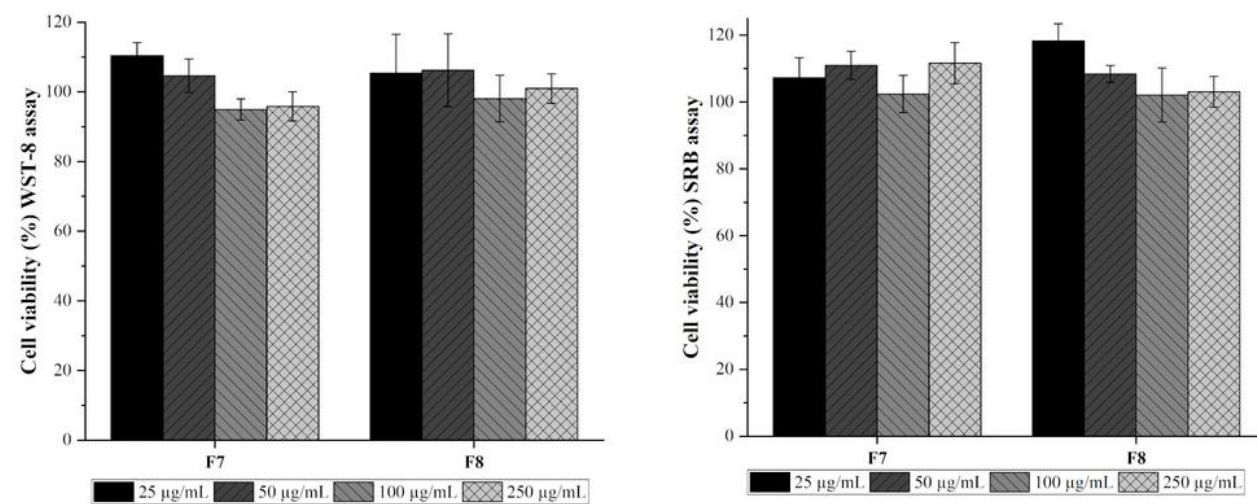
- Glycyrrhetic acid is an active compound present in licorice
- It showed anti-inflammatory and anti-oxidant activities when applied topically
- Ethosomes are known to have an increased permeation through stratum corneum
- Production using 3D printed microfluidic chip



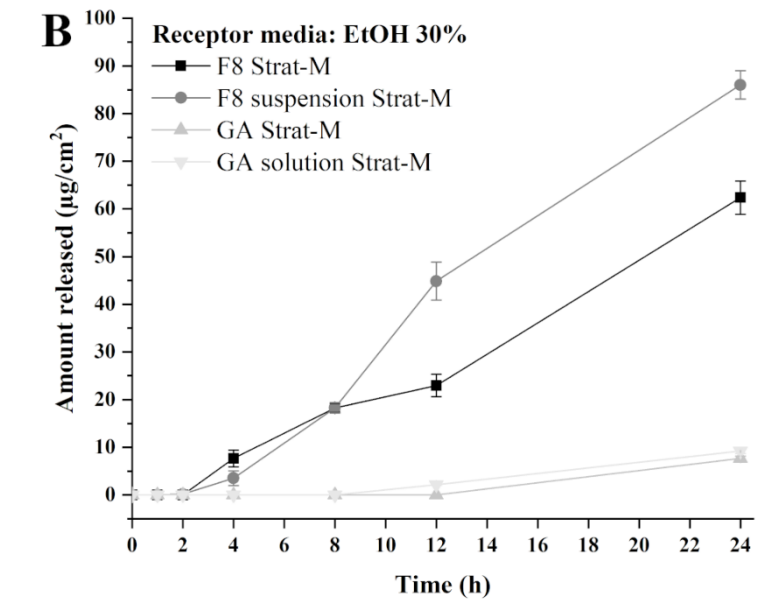
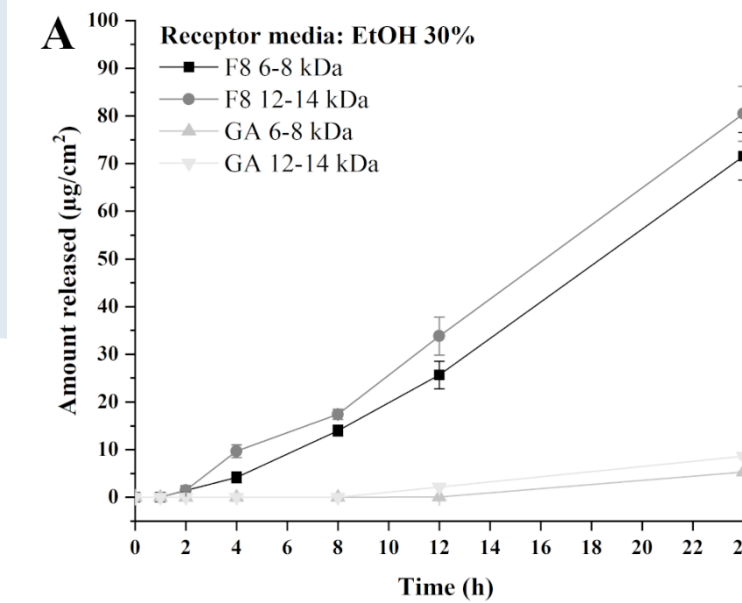
RESULTS



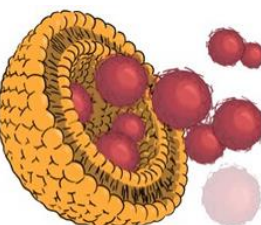
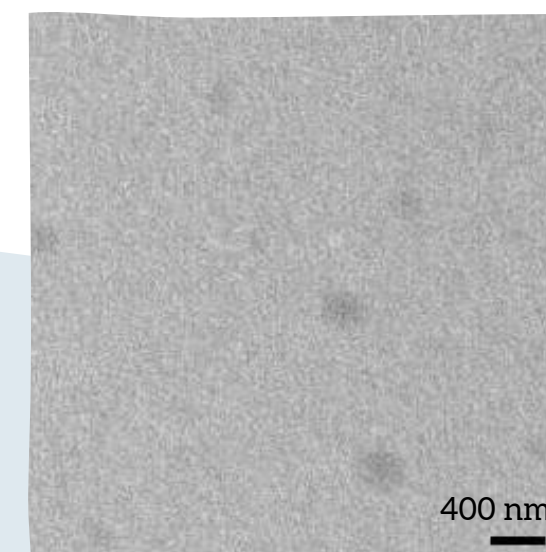
FTIR of GA, plain liposomes and GA-loaded liposomes



202 ± 5.2 nm
Narrow size distribution
Good stability over a period of 30 days
Encapsulation efficiency of 63.15 ± 2.2%,



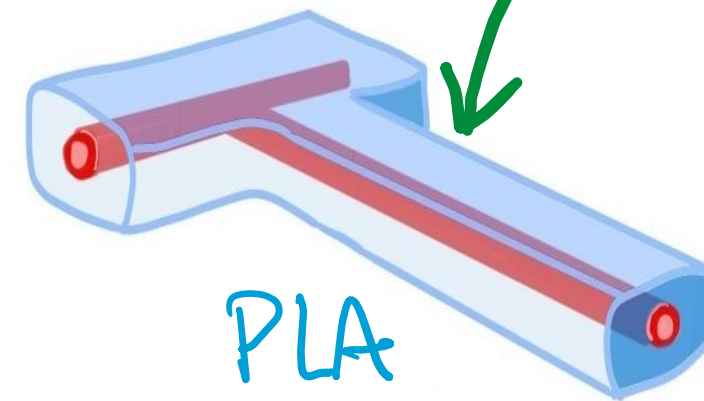
In vitro release and permeation studies using vertical diffusion cells (VDC) with Ethanol 30% (v/v) as receptor medium (A, B). In the graphs, the GA-loaded liposomal hydrogel, the GA-loaded liposomal suspension, saturated GA hydrogel and saturated GA solution are represented. Cellulose dialysis membranes with a different cut-off (A) and skin mimicking Strat-M® membranes (B) were used.



Cell viability of HaCaT cell line after incubation with unloaded liposomes and GA loaded liposomes at different concentrations for 24 h at 37 °C. The viability was determined by WST-8 and SRB assays

CONCLUSIONS

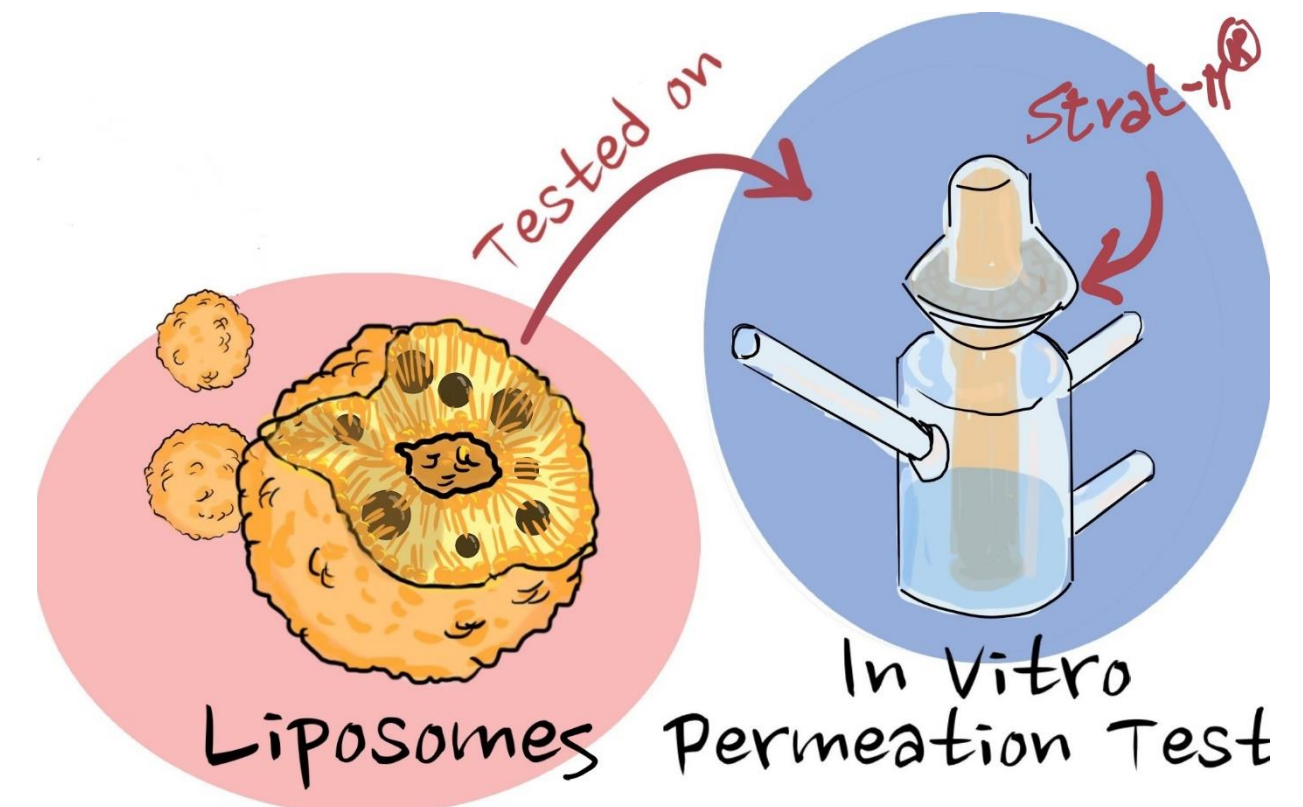
- Development of an ethanolic liposomal formulation encapsulating GA intended for topical administration with good physicochemical characteristics and an EE% of GA up to 63 %,
- Development of a biodegradable and cost-effective PLA 3D-printed microfluidic chip which represents an affordable microfluidic device with an easy fabrication, very low cost (13 g of PLA for a total cost of less than 1 US \$) and potential scalability for higher production rates (up to 900 mL/h).
- The liposomal hydrogel compared to GA-saturated hydrogel reached an almost 10x times higher drug release and an 8x times higher drug permeation across skin mimicking Strat-M® membranes that could represent a reproducible methodology using VDC, alternative to ex-vivo skin in diffusion studies.



PLA



< 1\$
Cost
Effective!



References: Pastorino G, Cornara L, Soares S, Rodrigues F Oliveira. M.B.P.P. 2018:2323-2339. Martins J P, Torrieri G, Santos H A. Expert Opin. Drug Deliv. 2018:469-479. Tiboni M, Benedetti S, Skouras A, Curzi G, Romano Perinelli D, Palmieri G F, Casettari L. Int. J. Pharm. 2020:584, 119436

Acknowledgements



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Thank you for your attention



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